

An autonomous molecular computer for logical control of gene expression

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Early biomolecular computer research focused on laboratory-scale, human-operated computers for complex computational problems^{1–7}. Recently, simple molecular-scale autonomous programmable computers were demonstrated^{8–15} allowing both input and output information to be in molecular form. Such computers, using biological molecules as input data and biologically active molecules as outputs, could produce a system for ‘logical’ control of biological processes. Here we describe an autonomous biomolecular computer that, at least *in vitro*, logically analyses the levels of messenger RNA species, and in response produces a molecule capable of affecting levels of gene expression. The computer operates at a concentration of close to a trillion computers per microlitre and consists of three programmable modules: a computation module, that is, a stochastic molecular automaton^{12–17}; an input module, by which specific mRNA levels or point mutations regulate software molecule concentrations, and hence automaton transition probabilities; and an output module, capable of controlled release of a short single-stranded DNA molecule. This approach might be applied *in vivo* to biochemical sensing, genetic engineering and even medical diagnosis and treatment. As a proof of principle we

programmed the computer to identify and analyse mRNA of disease-related genes^{18–22} associated with models of small-cell lung cancer and prostate cancer, and to produce a single-stranded DNA molecule modelled after an anticancer drug.

Taking our cue from the terminology of medical treatment, we consider that our molecular computer performs *in vitro* a computational version^{23,24} of ‘diagnosis’—the identification of a combination of mRNA molecules at specific levels, which in our example is a highly simplified model of cancer—and ‘therapy’—production of a biologically active molecule, which in our case is a drug-like single-stranded (ss)DNA with known anticancer activity (Fig. 1a). The computer operation is governed by a ‘diagnostic rule’ that encodes medical knowledge in simplified form (Fig. 1b). The left-hand side of the rule consists of a list of molecular indicators for a specific disease, and its right-hand side indicates a molecule to be released, which could be a drug for that disease. For example, the diagnostic rule for prostate cancer states²⁰ that if the genes *PPAP2B* and *GSTP1* are underexpressed and the genes *PIM1* and hepsin (*HPN*) are overexpressed then administer the ssDNA molecule GTTGGTATTGGACATG, which inhibits²⁵ the synthesis of the protein MDM2 by binding to its mRNA. The computer design is flexible in that any sufficiently long RNA molecule can function as a molecular indicator and any short ssDNA molecule, up to at least 21 nucleotides, can be administered.

The computation module is a molecular automaton^{12–15} (Supplementary Fig. S1) that processes such a rule as depicted in Fig. 1c. The automaton has two states: positive (Yes) and negative (No). The computation starts in the positive state and if it ends in that state we call the result ‘positive diagnosis’, otherwise it is called ‘negative diagnosis’. To facilitate rule processing by the automaton, the left-hand side of the diagnostic rule is represented as a string of symbolic indicators, or symbols for short, one for each molecular

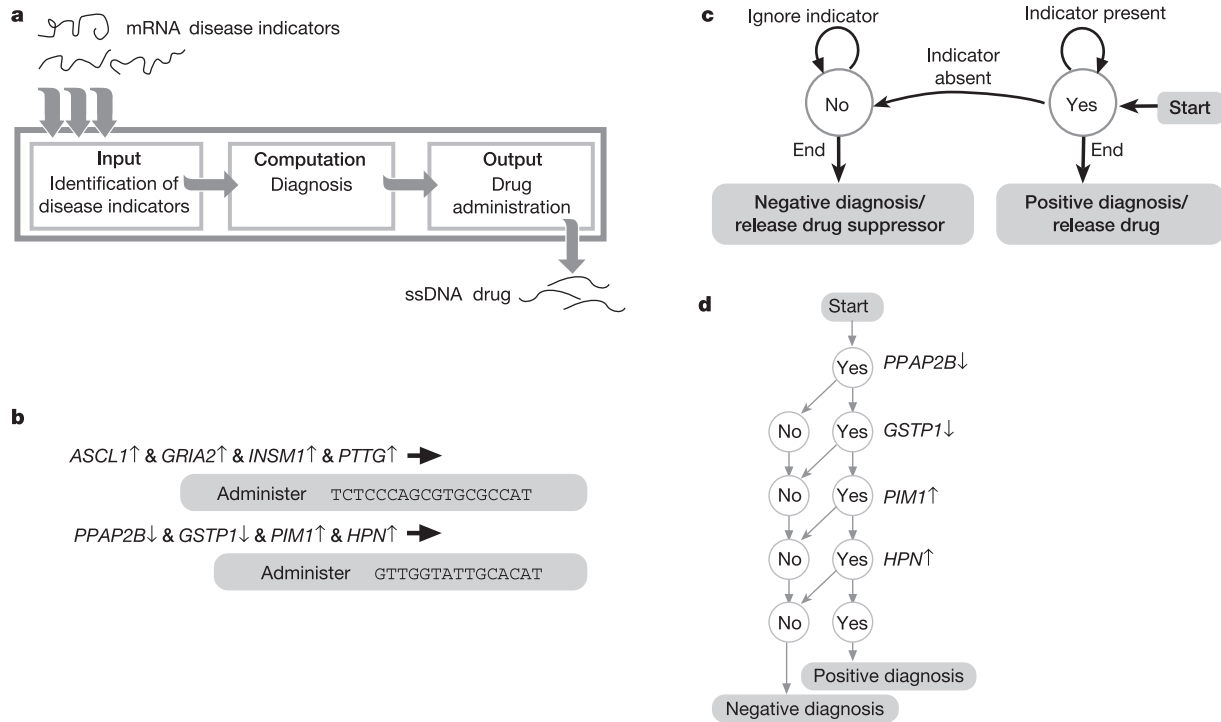


Figure 1 Logical design and logical operation of the molecular computer. **a**, Function and modular organization of the molecular computer. **b**, Example diagnostic rules for simplified models of SCLC¹⁹ and prostate cancer²⁰, indicating overexpression (↑) or underexpression (↓) of a disease-related gene. The first rule states that if genes *ASCL1*, *GRIA2*, *INSM1* and *PTTG1* are overexpressed then administer the ssDNA molecule

TCTCCAGCGTGCCAT (oblimersen), purported to be an antisense therapy drug for SCLC²⁷. The second rule states that if the genes *PPAP2B* and *GSTP1* are underexpressed and the genes *PIM1* and *HPN* are overexpressed then administer the ssDNA molecule GTTGGTATTGCACAT, purported to be a drug for prostate cancer²⁵. **c**, Transition diagram of the diagnostic automaton. **d**, The computation that diagnoses prostate cancer.

indicator. For example, the string for the prostate cancer rule is *PPAP2B* ↓ *GSTP1* ↓ *PIM1* ↑ *HPN* ↑. For each symbol the automaton has three types of transitions: positive (Yes → Yes), negative (Yes → No) and neutral (No → No). The automaton processes the string from left to right, one symbol at a time. When processing a symbol in the positive state, the computer takes the positive transition if it determines that the molecular indicator is present and the negative transition, changing to a negative state, otherwise. As the No → Yes transition is not allowed, once the automaton enters the negative state it can use only the neutral transition and thus remains in the negative state for the duration of the computation. The possible computation paths of the automaton processing the prostate cancer diagnostic rule are shown in Fig. 1d.

The molecular automaton is stochastic^{14,16} in that it has two competing transitions, positive and negative, for each symbol while in the positive state. A novel molecular mechanism, explained below, regulates the probability of each positive transition by the corresponding molecular indicator, so that the presence of the indicator increases the probability of a positive transition and decreases the probability of its competing negative transition, and vice versa if the indicator is absent. Because the confidence with which the presence or absence of an indicator can be determined is a continuous rather than a discrete parameter, so is the regulation of transition probabilities, the level of which is correlated with this confidence. The resulting stochastic behaviour of the automaton is governed by the confidence in the presence of each indicator, so that the probability of a positive diagnosis is the product of the probabilities of the positive transitions for each of the indicators processed (see Supplementary Note). By changing the ratio between positive and negative transitions for a particular indicator we can have fine control over the sensitivity of diagnosis to the presence of that indicator. We note our unusual use of automaton components: its formal input (the diagnostic rule to be processed) functions in our application like a program, and its formal program (the software molecules) functions in our application as part of the input module, detecting the presence of molecular indicators.

Instead of releasing an output molecule on positive diagnosis and doing nothing on negative diagnosis, we opted to release a biologically active molecule (for example, a drug) on positive diagnosis and its suppressor molecule on negative diagnosis. This allows fine control over the diagnosis confidence threshold beyond which an active drug is administered. Rather than using a single automaton for both tasks (which cannot be done with our current design), we implement this by using two types of automata: one that releases a drug molecule on positive diagnosis, and another that releases a drug-suppressor molecule on negative diagnosis. The ratio between the drug and drug-suppressor molecules released by a population of automata of these two types determines the final active drug concentration.

The molecular computer realizing this logical design consists of diagnostic molecules that encode diagnostic rules (Fig. 2a; see also Supplementary Fig. S2); transition molecules that realize automaton transitions and are regulated by molecular indicators (Fig. 2b; see also Supplementary Fig. S2); and hardware molecules (the restriction enzyme *FokI*) (Supplementary Fig. S1).

A diagnostic molecule has a diagnosis moiety and a drug-administration moiety (Fig. 2a; see also Supplementary Fig. S2). The diagnosis moiety realizes each symbol by a unique double-stranded (ds)DNA sequence 7 base pairs (bp) in length. For all symbol-encoding sequences, the first four nucleotides represent the symbol combined with the positive state and nucleotides three to six represent the symbol combined with the negative state. The automaton processes the diagnosis moiety one symbol at a time, determining whether the corresponding indicator is present (Fig. 1c). After all symbols are processed, the computation proceeds to processing the drug-administration moiety. There are two kinds of drug-administration moieties: drug release and drug-suppressor

release. Both consist of a double-stranded stem that protects a single-stranded loop containing the drug or the drug suppressor. The stem prevents interactions between the drug, drug suppressor and target mRNA. Its length (three symbols) was determined empirically (data not shown). After positive diagnosis, the stem of a drug-suppressor moiety is cleaved by special Yes-verification transitions and releases the drug; in the case of negative diagnosis it remains intact, protecting the drug (Fig. 2d). After negative diagnosis, the stem of the drug-suppressor release moiety is cleaved by special No-verification transitions and releases the drug suppressor; in the case of positive diagnosis it remains intact, protecting the drug suppressor (Fig. 2d).

The ratio between diagnostic molecules with a drug release moiety and those with a drug-suppressor release moiety need not necessarily be 1:1. For example, if the measured probability of positive diagnosis does not exceed 50% owing to computer design limitations, combining the two types of molecules at a ratio of 2:1 will render an active drug for 1/6 of the computations. Inverting this ratio can compensate for false-positive diagnosis by suppressing drug release below any desired threshold.

The molecules released in our experiments are the antisense ssDNA for MDM2 (GTTGGTATTGCACAT)²⁵ and the ssDNA molecule having the same sequence as the drug Vitravene²⁶ (GCGTTTGCTCTTCTTGTGCG) (Supplementary Data). Similar molecules, such as oblimersen²⁷ (TCTCCAGCGTGCGCCAT), could similarly be released. The same method can be used to release short RNA molecules.

For each symbolic indicator, a pair of competing transition molecules (Fig. 2b; see also Supplementary Fig. S2) perform the corresponding molecular indicator verification. Presence of a molecular indicator entails high concentration of the positive transition molecule and low concentration of its competing negative transition molecule and vice versa. This regulation is accomplished by sequence-specific interaction between the indicator and a partially single-stranded fragment of a transition molecule, as follows. A positive transition checking for overexpression is activated by a high concentration of its corresponding mRNA, whereas a positive transition checking for underexpression is inactivated by a high concentration of its corresponding mRNA. The corresponding negative transitions are oppositely regulated by a similar mechanism (Fig. 2b; see also Supplementary Fig. S2). A similar mechanism allows for transition regulation by point mutation (Supplementary Fig. S2). The logical switch between configurations of the transition molecules is similar to the state that switching a DNA fuel molecule causes in a DNA nanoactuator²⁸. We also note an alternative approach to sensing biochemical signals, known as ‘chemical logic gates’^{29,30}.

The computer consists of three molecular modules—input (Fig. 2b), computation (Fig. 2a) and output (Fig. 2d)—that interact with the disease-related molecules and with each other via a complex network (Fig. 2). Each molecular computer autonomously performs one diagnosis and drug administration task; multiple tasks can be performed by multiple computers that operate in parallel within the same environment without mutual interference, while sharing the hardware molecules and potentially sharing some or all of the software molecules. All pairs of transition molecules are regulated simultaneously by their respective indicators (Fig. 2b) and perform a stochastic computation over diagnostic molecules to administer a drug upon diagnosis (Fig. 2d).

To provide a successful implementation, the computer must be robust both to imprecision of molecular components and to variations in external parameters. This is achieved by three mechanisms. First, imprecision in transition regulation may be compensated by variation in the relative amounts of the active and inactive transition molecules and by addition of excess ssDNA oligonucleotides that form these transitions. Second, changes in the absolute level at which a molecular indicator should be positively detected

are compensated for by a similar change in absolute concentration of the transition molecules (Supplementary Fig. S3). Third, false-positive or false-negative diagnoses may be compensated for as explained above.

The operation of the computer modules was verified separately and jointly: transition regulation by the input module (Fig. 2b; see also Supplementary Fig. S2) was verified independently (Fig. 3a; see also Supplementary Fig. S3); the input and computation modules

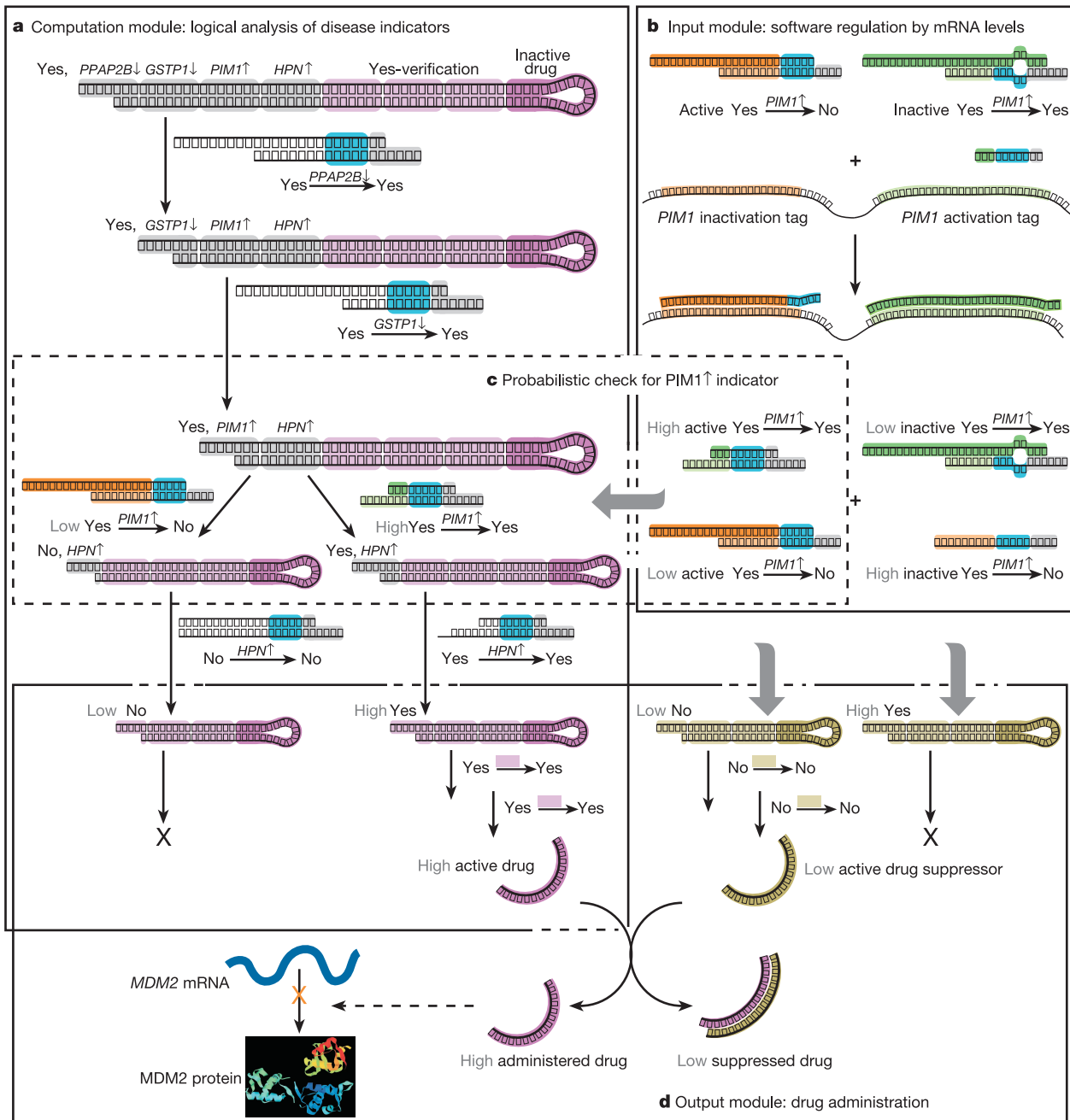


Figure 2 Operation of the molecular computer. The complete sequences for all molecules shown are given in the Supplementary Methods. **a**, Part of the computation path for the diagnostic molecule for prostate cancer with all molecular indicators present, ending in drug release. The initial diagnostic molecule consists of a diagnosis moiety (grey) that encodes the left-hand side of the diagnostic rule (Fig. 1b) and a drug-administration moiety (light purple) incorporating an inactive drug loop (dark purple). At each computation step, the prevailing transition is shown, except for the processing of the symbol $PIM1 \uparrow$, for which details of the stochastic choice, accomplished by a regulated pair of competing transition molecules, are shown (dashed box, see **c**). **b**, Regulation of the two transitions for $PIM1 \uparrow$ by sub-sequences (tags) of overexpressed $PIM1$ mRNA, resulting in a relatively high level of the $Yes \xrightarrow{PIM1 \uparrow} Yes$ transition molecules and low level of the $Yes \xrightarrow{PIM1 \uparrow} No$ molecules. Each transition molecule contains regulation (green, orange) and computation (blue, grey) fragments. The ‘inactivation tag’ of $PIM1$ mRNA (light

orange) displaces the 5' → 3' strand of the transition molecule $Yes \xrightarrow{PIM1 \uparrow} No$ and destroys its computation fragment. The ‘activation tag’ of $PIM1$ mRNA (light green) activates the transition molecule $Yes \xrightarrow{PIM1 \uparrow} Yes$. Initially, a protecting oligonucleotide (green) partially hybridizes to the 3' → 5' strand of the transition molecule and blocks the correct annealing of its 5' → 3' strand. The ‘activation tag’ displaces the protecting strand, allowing such annealing and rendering an active $Yes \xrightarrow{PIM1 \uparrow} Yes$ transition. Ideally, one $PIM1$ mRNA molecule inactivates one $Yes \xrightarrow{PIM1 \uparrow} No$ and activates one $Yes \xrightarrow{PIM1 \uparrow} Yes$ transition molecule. **c**, Stochastic processing of the symbol $PIM1 \uparrow$ by a regulated pair of competing transition molecules. The probability of a $Yes \rightarrow Yes$ transition is high, resulting in a high level of diagnostic molecules in the state Yes and a low level in state No. **d**, Combining computation results for both types of diagnostic molecules, both with high Yes and low No final states, results in high release of drug and low release of drug suppressor, and hence in the administration of the drug.

performing transition regulation and diagnostic computation (Fig. 2a) were verified together (Fig. 3b, c); finally, the input, computation and output modules were combined to perform transition regulation, diagnostic computation and drug administration (Figs 2d and 4a).

The regulation of transition molecules by mRNA (Fig. 2b) was demonstrated (Fig. 3a) with an mRNA transcript of about 1,900 nucleotides as an example indicator. A correlation between the mRNA level and the probability of positive diagnosis, effected by a pair of transitions regulated to check for underexpression, is shown in Supplementary Fig. S3. Regulation by mutation was confirmed with a ssDNA model simulating a point substitution representing a small-cell lung cancer (SCLC)-related mutation in the gene

encoding p53 (ref. 21) (Supplementary Fig. S3). We expect to extend this approach to detect insertion and deletion mutations.

In diagnosing a disease model with multiple indicators we used ssDNA oligonucleotides to represent disease-related mRNA and two concentrations to represent mRNA levels: 0 μ M for low level and 3 μ M for high level. Transition regulation can be adjusted by changing the absolute concentration of competing transitions to distinguish between no mRNA and mRNA at concentrations as low as 100 nM, which represent about 50 mRNA copies per mammalian cell. In our experiments we used up to four molecular indicators, although the specific symbol encoding used can provide up to eight indicators.

We tested the input and computation modules of the computer

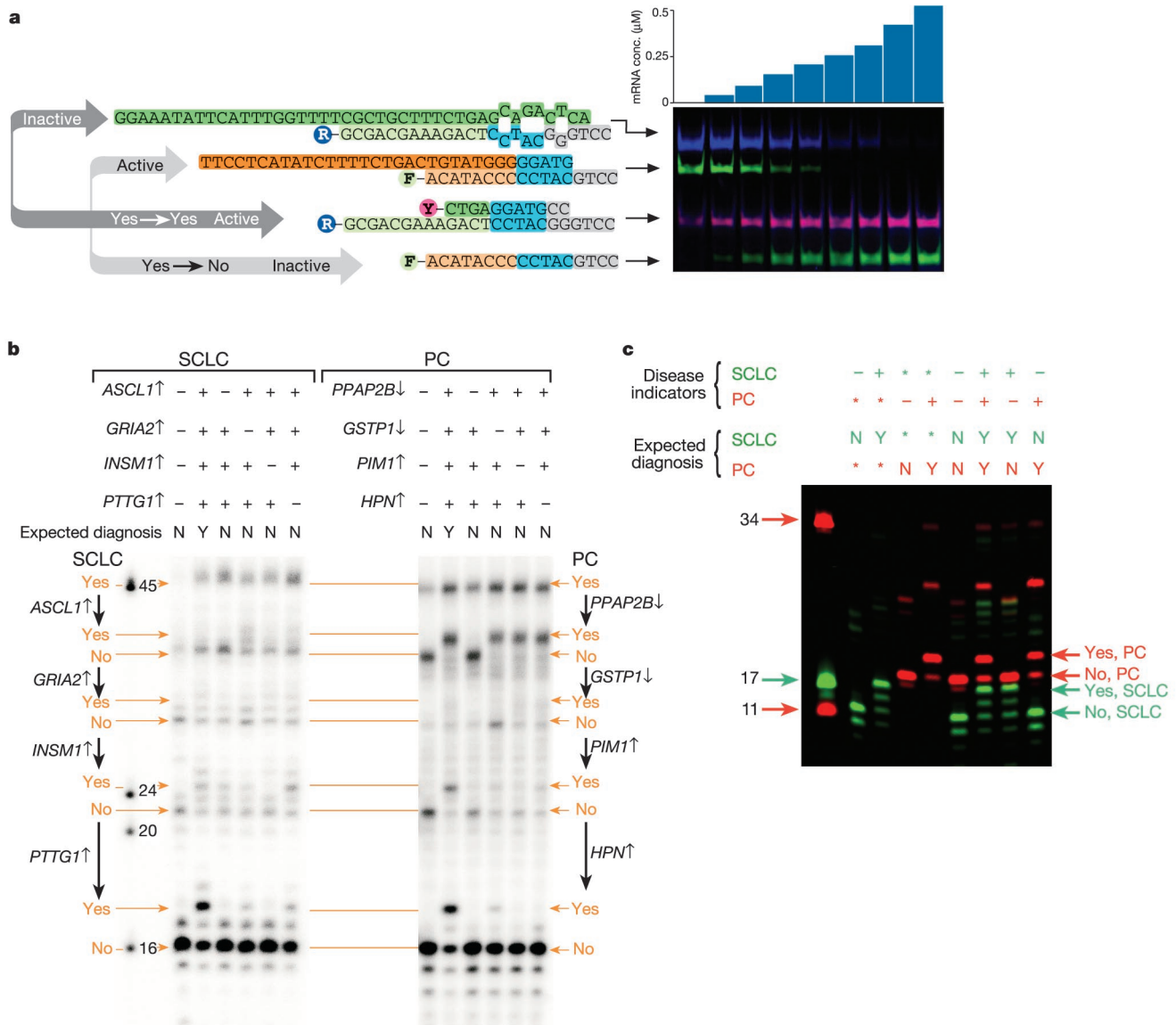


Figure 3 Experimental demonstration of diagnosis. **a**, Regulation of competing transitions by pTRI-Xef mRNA (bar chart) representing an example molecular indicator. The gel shows positive (blue and pink) and negative (green) transition, each in both active and inactive states, the relative concentration of which is regulated by mRNA concentration. F, carboxyfluorescein; R, tetramethyl rhodamine; Y, Cy5. **b**, Validation of the diagnostic automata with the diagnostic rules for SCLC and prostate cancer (PC). Each lane shows the result of a diagnostic computation for the indicated combination of molecular indicators. The location of the initial, intermediate and output molecules is indicated on

the left and a predicted trace of the diagnostic computations is shown on both sides. Some, but not all, intermediates are visible owing to their incomplete processing by the automaton. **c**, Parallel diagnosis of two disease models by two diagnostic automata. The diagnosed disease model contains the two indicators of SCLC checked by the diagnostic string *PTTG1* \uparrow *CDKN2A* \uparrow and the two indicators of prostate cancer checked by the string *PIM1* \uparrow *HPN* \uparrow . The gene *CDKN2A* is another indicator for SCLC when overexpressed, and it is used here for technical reasons. The presence of indicators for each disease model as well as the expected diagnostic output by each automaton are indicated above the lanes.

on molecular models of SCLC and prostate cancer with diagnostic automata for the diagnostic rules shown in Fig. 1b. The diagnostic output was identified by the size of an inert fragment released on completion of the computation. Each automaton reaches positive diagnosis with significant probability only when all four molecular indicators are present (Fig. 3b). The false-negative diagnosis is due to limitations in the design of the transition molecules, but can be compensated for during drug administration (Fig. 4c) as discussed above. We also confirmed the selectivity of the diagnostic process by testing the two diagnostic automata in mixed conditions, in which molecular indicators for none, one, or both disease models are present (Supplementary Fig. S4). In all cases a positive diagnosis was made with significant probability by a diagnostic automaton only when all molecular indicators were present. We confirmed the possibility of parallel diagnosis of multiple disease models by running together two diagnostic automata under various compositions of molecular indicators (Fig. 3c). Indeed, each automaton performed its diagnosis correctly, irrespective of the computation performed by the other automaton.

Drug administration is demonstrated for the prostate cancer disease model (Fig. 4). We constructed the drug-release diagnostic molecule for *PPAP2B* ↓ *GSTP1* ↓ *PIM1* ↑ *HPN* ↑ (Fig. 2a) and tested the extent of active drug release for different diagnostic outcomes,

effected by varying amounts of ssDNA representing *HPN* mRNA and, in a separate experiment, an example mRNA that substitutes for *GSTP1* mRNA. The presence of other indicators was modelled by appropriately formed positive transitions. We show that the amount of active drug increases with the confidence in a positive diagnosis (Fig. 4a). We demonstrate the concept of drug regulation by a drug suppressor using diagnostic molecules for *PPAP2B* ↓ *GSTP1* ↓ with drug release and drug-suppressor release moieties. We show that the prevailing species is the active drug for high probability values, a drug/drug-suppressor hybrid for intermediate probability values, and an active drug suppressor for low probability values (Fig. 4b). We also demonstrate the ability to control the relative amounts of drug and drug suppressor for the 1:1 ratio of positive and negative diagnosis (Fig. 4c). The result demonstrates the robustness of our proposed compensation mechanism and illustrates how multiple degrees of freedom of the system allow it to overcome imperfections in its components.

Our work demonstrates a robust and flexible molecular computer capable of logical analysis of mRNA disease indicators *in vitro* and the controlled administration of biologically active ssDNA molecules, including drugs. The modularity of the design facilitates improving each computer component independently. For example, computer regulation by other biological molecules such as proteins,

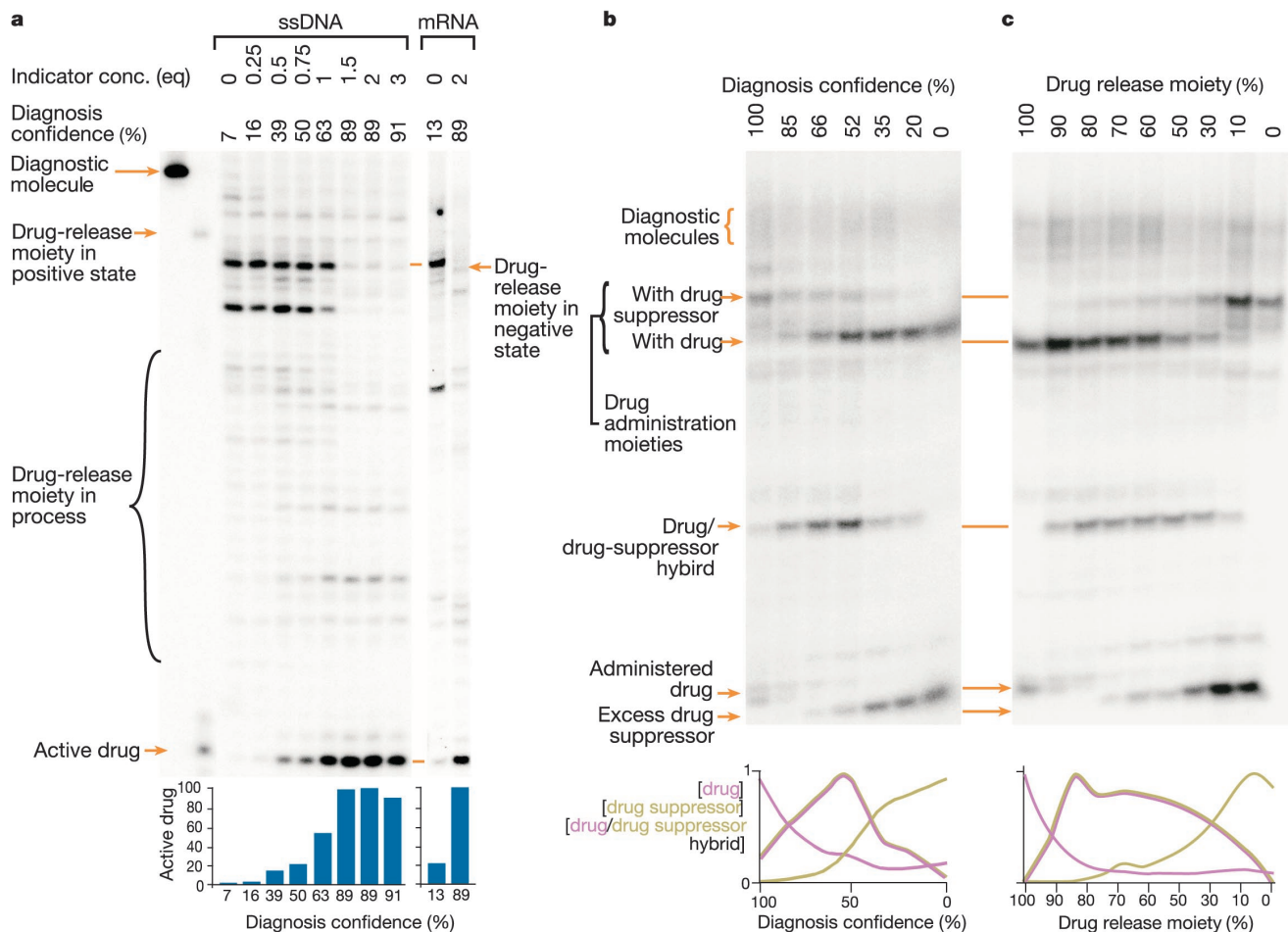


Figure 4 Experimental demonstration of drug administration. **a**, Release of an active drug by a drug-release *PPAP2B* ↓ *GSTP1* ↓ *PIM1* ↑ *HPN* ↑ diagnostic molecule, showing absolute amount of the active drug versus positive diagnosis probability. **b**, Different diagnostic outcomes are modelled using active transition molecules with a mixture of equal amounts of the drug-release and drug-suppressor release moieties for the diagnostic string *PPAP2B* ↓ *GSTP1* ↓. Each lane shows the distribution of drug-

administration moieties, active drug, excess drug suppressor and drug/drug-suppressor hybrid, as indicated. **c**, Variation in the distribution of active drug, excess drug suppressor and drug/drug-suppressor hybrid for a given diagnostic outcome and for varying relative amounts of drug release and drug-suppressor release diagnostic moiety. *y*-axis units in **b** and **c** are arbitrary units.

the output of other biologically active molecules such as RNA interference, and *in vivo* operation can all be explored simultaneously and independently. □

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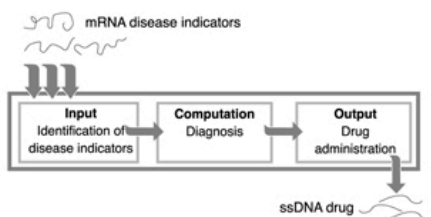
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a Modular design of the molecular computer

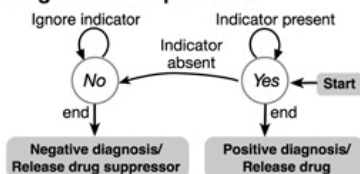


b Diagnostic rules for small-cell lung cancer and for prostate cancer

ASCL1↑ & GRIA2↑ & INSM1↑ & PTTG↑ →
 Administer TCTCCAGCGTGCACAT

PPAP2B↓ & GSTP1↓ & PIM1↑ & HEP SIN↑ →
 Administer GTTGGTATTGCACAT

c Diagnostic rule processor



d Processing prostate cancer diagnostic rule

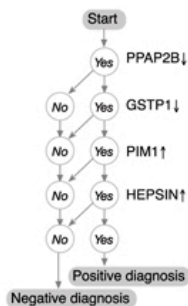


Fig. 1:

Logical design of the Molecular computer:

- a) Each computer consists of three, semi-independent modules. To a certain extent, changes may be made in one of the modules without affecting the other two.
- b) The input module checks for the presence or absence of up to eight indicators for disease - (upper left letters) The computation module processes this data and performs diagnosis by giving the "certainty level" for the presence of the disease. The output module can now act accordingly by releasing a drug (letters in gray) to fit the diagnosis.
- c) A graphic scheme of the computer (automaton) design. If a cancer marker is detected, the automaton will stay in the initial "Yes" state. If, however, one marker is absent, the automaton will move to the "No" state, with no possible return to the "Yes" state. The computer then "examines" the final state, making its diagnosis, and releases a drug or drug suppressor

accordingly.

- d) A tree-diagram of a computation process. Diagnosis proceeds one marker at a time. Failure to detect a marker at any point will switch the system from "Yes" to "No," and the computation module remains in this state, irrespective of the detection of further markers.

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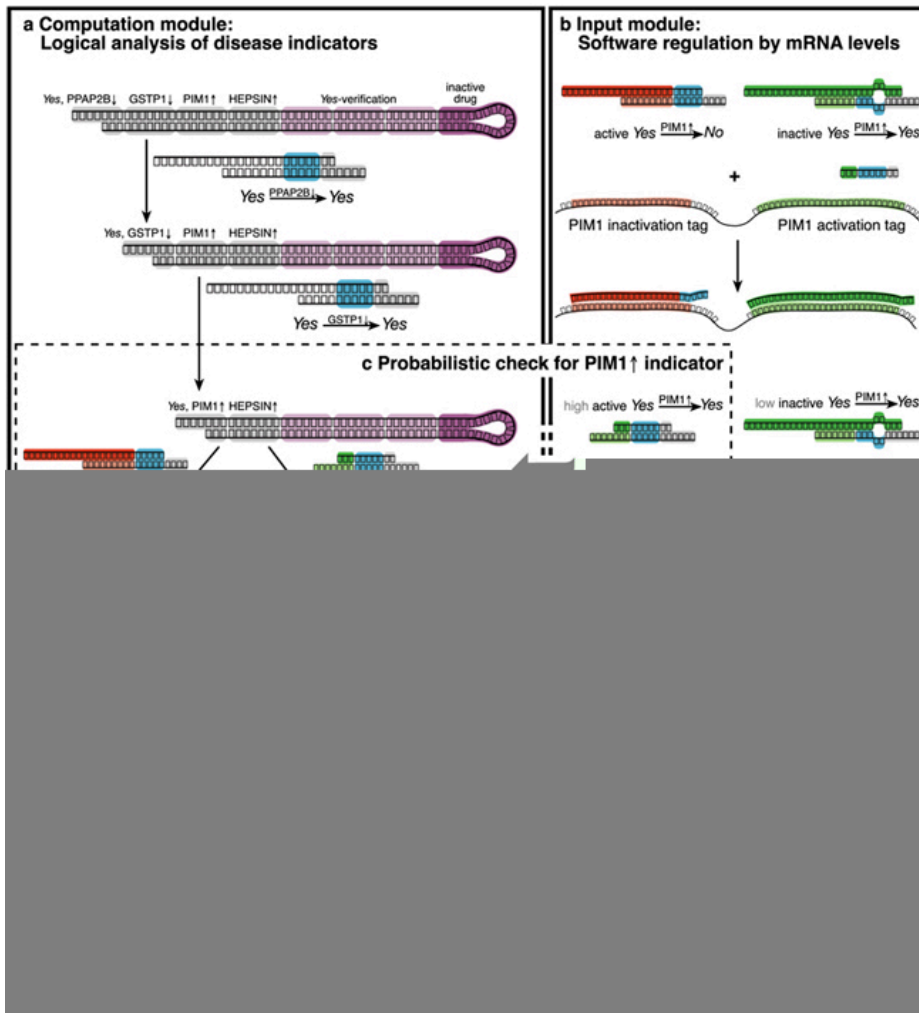


Fig. 2:

Schematic layout of the molecular computer's operation:

- a) The overall process: The gray section is made up of four smaller segments, or symbols, each of which checks for a different disease marker. The middle module (purple or beige) verifies the final diagnosis, and facilitates the release of either a drug or drug suppressor (darker colored loop at the end).
- b) As each test takes place in the diagnostic process, there are three possible transitions to the next stage: Yes \rightarrow Yes (positive), Yes \rightarrow No (negative) or No \rightarrow No (neutral). Transition molecules carry information about the presence or absence of disease markers. In our example, the transition molecules respond to the presence of an mRNA molecule. To detect overexpression of the mRNA, we designed the transition process such that positive detection of the mRNA molecule will build the positive transition and annul the negative transition.
- c) Computation continues, one indicator at a time. The first two markers have evoked strong positive (Yes \rightarrow Yes) transitions, but levels of the third marker are lower. Therefore, there will be more negative transitions, though the overall number of positive transitions remains high relative to the negative's. The computations proceed to the next indicator, though modules in the "No" state must remain that way.
- d) Up till now, diagnosis has proceeded in an identical manner for both drug and suppressor-carrying computers. At the end of the diagnostic computation process, one of two final transitions takes place. One transition molecule detects only the modules that carry the drug and are in the "Yes" state (purple), while the second detects those with the suppressor that are in the "No" state (beige). These molecules then snip off the looped end of the computer molecule, thereby activating it. Suppressors will bind (by hybridization) with drugs in equal amounts to cancel them out, but, if there are more positive than negative results, some drug molecules will remain.

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Molecular components and computational step of a molecular automaton

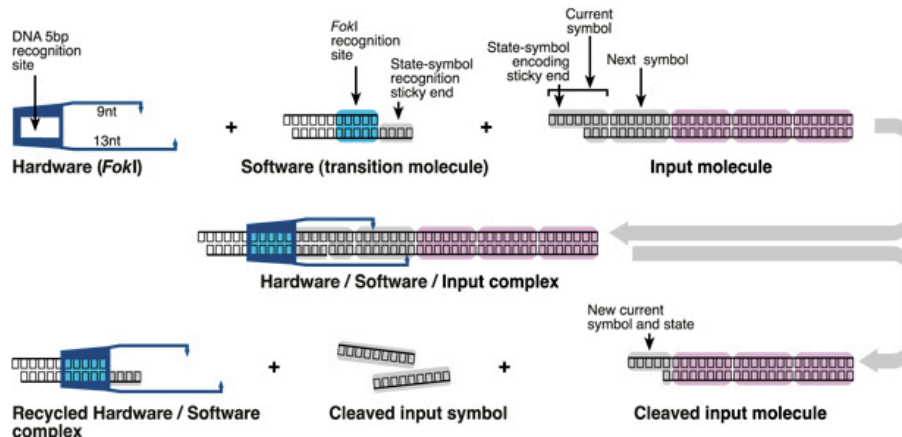
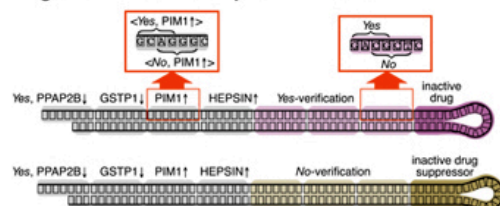
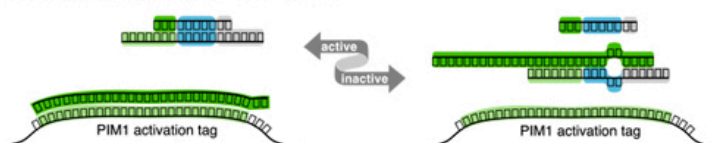


Figure S1. Architecture of the molecular finite automaton, featuring its input, software and hardware components. The current state of the computation is represented by a partially cleaved symbol-encoding dsDNA segment that exposes a four-nucleotide "sticky end" at a state-specific location. The cleavage is accomplished by the FokI hardware enzyme that recognizes the double-stranded DNA sequence GGATG and cleaves its substrate 9 and 13 nucleotides away from the recognition site in 5'_'3' and 3'_'5' strands, respectively. The transition molecule recognizes a particular state-symbol sticky end and directs the FokI bound to it to cleave within the next symbol at a precise location to expose the next state-symbol combination, thus realizing transition between states. The software molecule is recycled and the cleaved symbol is scattered. We note our unusual use of automaton components: its formal input, the diagnostic rule to be processed, functions in our application like a program, and its formal program, the software molecules, function in our application as the input mechanism, detecting the presence of molecular indicators.

a Diagnostic molecules for prostate cancer



b Transition molecule for Yes^{PIM1} → Yes



c Transition molecule for Yes^{PIM1} → No



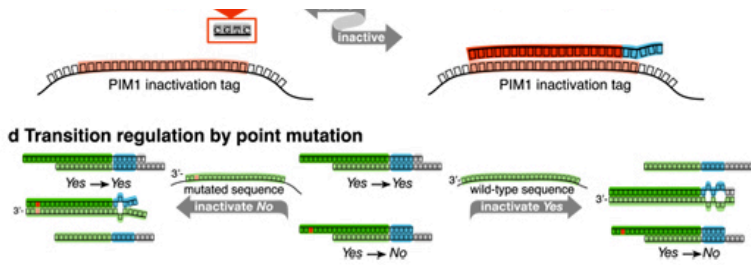


Figure S2. Molecular components of the computer. a, Diagnostic molecules for prostate cancer. The diagnosis moiety (gray) implements the diagnosis component of a diagnosis and therapy rule and it consists of 7-bp sequences encoding the symbols for the molecular indicators. Following it are either a drug release moiety (purple) or a drug-suppressor release moiety (brown), consisting of a ssDNA that loops on itself to form a sequence encoding three diagnostic verification symbols (light purple/light brown) followed by a drug loop (purple) or a drug-suppressor loop (brown). For all symbol-representing sequences, the first four nucleotides of the sequence represent the symbol combined with state Yes, while nucleotides three to six represents the symbol combined with the state No. Example symbol encodings and state-symbol sticky ends are enlarged in red frames. b, c, Pair of competing transition molecules regulated by PIM1 mRNA, each containing a regulation (green, red) and a computation (blue, gray) fragment. The computation fragment consists of the double-stranded recognition site of the hardware enzyme FokI (blue), a single-stranded sticky end (gray) that recognizes a particular state-symbol combination of the diagnostic molecule, and possibly a 2-bp spacer (gray) between the two. A spacer of 2 bp effects a Yes→Yes transition while a zero-length spacer effects a Yes→No transition. The regulation fragment of a transition molecule enables its regulation by a nucleic-acid-based molecular indicator, which may activate (green) or deactivate (red) the transition when in high concentration. The transition molecule Yes→No (c) is inactivated by a subsequence of the PIM1 mRNA indicator ("inactivation tag") via its binding to the single-stranded overhang of the regulation fragment of the transition molecule followed by strand exchange due to higher stability of the mRNA-deactivation-tag/transition-sense-strand hybrid relative to the normal transition molecule hybrid. The transition molecule Yes→Yes (b) is activated by high concentration of PIM1 mRNA. In its absence, formation of the transition molecule is prevented by a third "protecting" oligonucleotide (green) that partially hybridizes to the antisense strand and forms a complex that is more stable than the active transition molecule. The protecting strand is also complementary to a subsequence of PIM1 mRNA ("activation tag", light green). Activation tag of PIM1 mRNA triggers a strand exchange process that decouples the protecting strand from the antisense strand of the transition molecule and allows it to hybridize with the sense strand to form an active Yes Yes transition. In an idealized regulation process one PIM1 mRNA molecule inactivates one Yes→No and activates one Yes Yes transition molecule. d, Pair of transition molecules regulated by mRNA point mutation. The positive transition has a regulation fragment complementary to the wild-type mRNA while the corresponding regulation fragment of the negative transition is complementary to the mutated mRNA. The positive transition is preferentially inactivated by the wild-type mRNA whereas the negative transition is inactivated by the mutated mRNA.

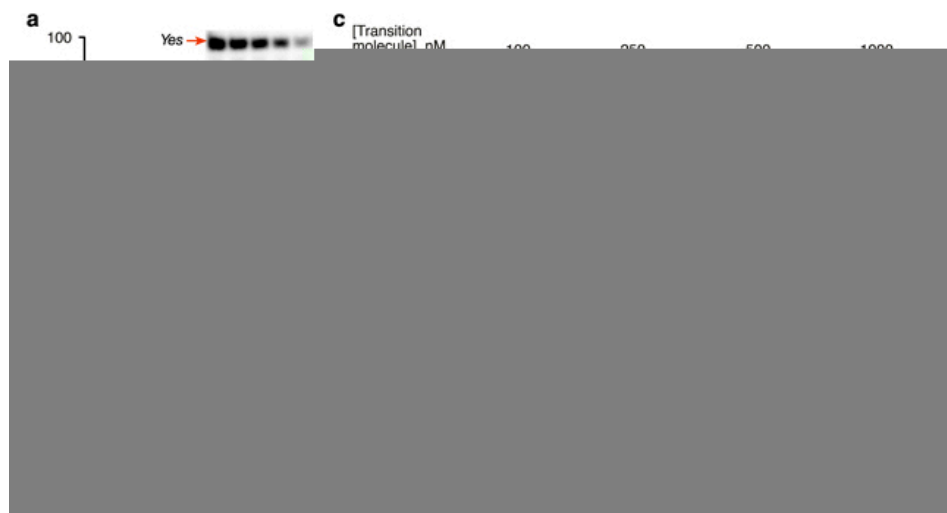


Figure S3. a, Calibration curve showing regulation of probability of Yes output state in a single-step computation by a pTRI-Xef generic mRNA indicator. Experimental data used to calculate the probabilities is shown in the insert. b, Regulation by point mutation by mixtures of model ssDNA oligonucleotides representing different ratios of mRNA of wild-type and of mutated genes. c-e, Adjusting confidence in a positive diagnosis for various concentrations of the transition indicator by adjusting the absolute concentrations of the transition molecules. The gel (c) visualizes the increase in probability of Yes diagnostic output with increasing concentrations of INSM1 ssDNA model (over-expressed in the disease) for different concentrations of active and inactive transition molecules. (d) Display of the transition probabilities derived from the measured intensities of the Yes and No bands, highlighting the change in the No/Yes crossover point as a function of transition molecule concentration. (e) Plots this function.

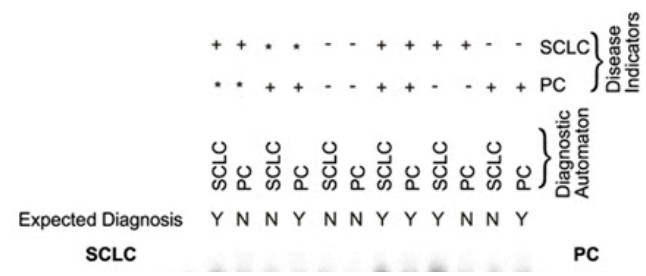




Figure S4. Selectivity of the diagnostic automata for their disease models. Each pair of lanes is a particular combination of molecular indicators indicated in the figure and is diagnosed separately by the automata for SCLC (left lane) and PC (right lane). + indicates presence of disease indicators, - indicates a normal condition, and * indicates absence of disease-related molecules. Expected outcome of the diagnosis is indicated above each lane.

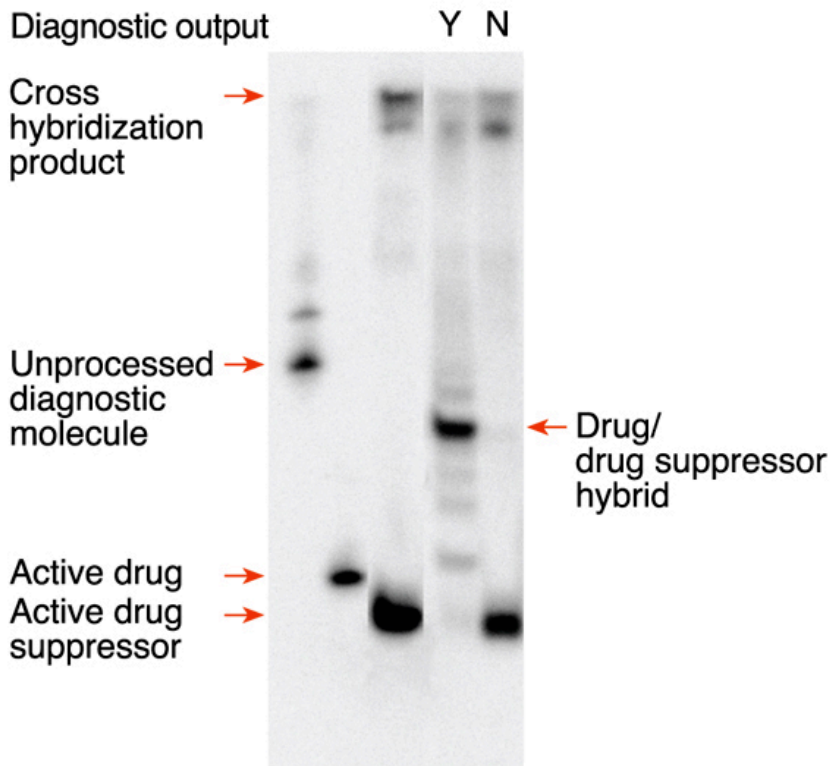


Figure S5. Release of the approved antisense drug. The two left-most lanes show, separately, a labelled diagnostic molecule and a labelled drug molecule. The third lane contains a non-labelled diagnostic molecule together with a labelled drug suppressor, incubated in NEB4 buffer. The fourth lane shows the release of an active Vitravene® drug upon positive diagnosis as visualized by the labelled drug suppressor probe. The right-most lane demonstrates that the active drug is not released upon negative diagnosis.

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